

## The effect of melatonin on apoptosis in hepatocellular carcinoma cell line: An experimental study

Sara Elsaadawy, Reda Elbadawy, Badawy Abd Elaziz, Heba Elkareem, Ghadeer Mosaad



### Summary

**Introduction:** Apoptosis resistance in Hepatocellular Carcinoma (HCC) is considered as one of the most important factors involved in hepatocarcinogenesis and its progression. Recent experimental research of melatonin highlights its anticancer function through its antioxidant effect, induction of apoptosis and cell cycle arrest, inhibition of metastasis, and suppression of angiogenesis. The aim of our study is to explore the effect of melatonin on casp 8 mRNA expression and Alpha-fetoprotein (AFP) mRNA expression in HepG2 HCC cell line by using real time PCR technique to proof its apoptotic effect. **Methods:** HepG2 cells were treated with 75mg melatonin then AFP mRNA expression and casp8 expression is investigated by RT PCR. **Results:** Melatonin significantly increased caspase 8 expression. The relative expression was 2.51, 2.48 and 1.89 respectively as compared to control ( $P < 0.001$ ). While the expression of AFP was down regulated by melatonin administration. The relative expression of AFP was 0.051, 0.052 and 0.046 respectively as compared to control ( $P < 0.001$ ). **Conclusion:** melatonin decreases AFP gene expression and stimulates apoptosis via up regulation of caspase 8 expression in HCC. Melatonin can be considered as promising adjuvant for chemotherapy in HCC.

**Keywords:** Alpha fetoprotein (AFP), Hepatocellular Carcinoma (HCC), Apoptosis, melatonin

Medical Journal of Viral Hepatitis

(MJVH) 2019; 4 (2) - pp. 75-79

Received: 3/11/2018

Revised: 7/2/2019

Accepted: 29/3/2019

Published Online: 25/4/2019

(Sara Elsaadawy) Resident of Gastroenterology & Hepatology dept, Damietta Cardiology and Gastroenterology center Egypt.

(Reda Elbadawy, Badawy Abd Elaziz, Ghadeer Mosaad) Hepatology, Gastroenterology & Infectious Diseases dept., Benha Univ., Egypt.

(Heba Elkareem) Medical Biochemistry dept., Faculty of Medicine, Benha Univ., Egypt.

\* CA: Sara Elsaadawy

drsara221@gmail.com

### Introduction

HCC represents a serious public health issue in Egypt, where liver cancer counts for 11.75% of the malignancies of the digestive organs and 1.68% of the total malignancies. HCC forms 70.48% of all liver tumors among Egyptians and it is considered the main complication of cirrhosis<sup>1</sup>. A number of strategies such as surgery, chemotherapy, molecular targeted therapy, radiation, and liver transplantation have been utilized for the treatment of HCC; however, most of the therapeutic treatments have a low response rate and are accompanied with significant side effects. Moreover, the high fatality rate of HCC has not yet improved, primarily because of uncontrolled metastasis, liver failure, and its resistance to therapeutic agents. To overcome these clinical shortcomings, genetic therapy has been considered as a new therapeutic approach for HCC<sup>2</sup>. AFP has become a target for liver cancer

immunotherapy<sup>3</sup>. It is a diagnostic marker for HCC. A direct relationship between poor prognosis and the concentration of serum AFP has been observed<sup>4</sup>. Apoptosis is one of the main mechanisms implicated in cell death, and its inactivation contributes to tumour progression and chemotherapy resistance<sup>5</sup>. The intrinsic pathway involves the mitochondrial release of cytochrome c and consequently procaspases activation. And this process is regulated by the Bcl-2 family of proteins<sup>6</sup>. The cascade of apoptosis signal transduction begins by the action of initiator caspases that are recruited and activated by autocatalytic processing. Among these caspases, caspase 8 and 9 are the main initiators of programmed cell death; caspase 8 is stimulated in response to extrinsic death ligands, while caspase 9 is necessary for the activation of executor caspase<sup>7</sup>. Targeting both the intrinsic and

extrinsic pathways reduces the growth of different tumour types, and a large number of studies demonstrate that different drugs alone or in combination enhance apoptosis in cancerous cells, including HCC<sup>8</sup>. A large number of studies have demonstrated that Melatonin has important oncostatic properties. It inhibits cell proliferation in several cancer cell lines including human B-lymphoma cells, human myeloid leukemia cells and human neuroblastoma cancer cells<sup>9</sup>. Melatonin decreased the growth rates of tumors in vivo both in transplantable animal model and the animal model induced by the administration of carcinogens<sup>10</sup>. This study aimed to investigate the effects of melatonin on apoptosis and AFP mRNA expression in HepG2 cell line by reverse transcription polymerase chain reaction (RT-PCR) in order to develop a new adjuvant therapy for HCC.

### Materials and methods

Melatonin (M.W 232.28 g/mol) was purchased from Sigma chemicals (Sigma Chemical Co., St. Louis, MO, USA) Melatonin was provided as 5 mg powder. The powder was dissolved in ethanol 95% forming 1 ml stock solution then it was diluted as 50 mol/L solution.

### Cell culture

Hepatic cancer cell line (HepG-2) was kindly supplied from tissue culture department VACSERA - Egypt. The cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% heat-inactivated fetal bovine serum (FBS), 100 IU/ml penicillin, and 100 mg/ml streptomycin in a humidified atmosphere of 95% air and 5% CO<sub>2</sub> at 37 °C. When cells became >80% confluent, cells were seeded into 96-well plates for 24 h in DMEM culture medium before they were treated with 75mg melatonin for 48 h before RT-PCR assay.

### Assessment of casp 8 and AFP gene expression by real-time quantitative reverse transcription polymerase chain reaction

Total RNA was isolated using the RNeasy extraction kit (Qiagen- Germany) according to the manufacturer's protocol. The RNA concentration was measured using nanodrop (Biowave II Germany). The RNA was reverse transcribed using the iScript cDNA Synthesis Kit (Bio-Rad, USA). Quantitative PCR was

performed using the Ready Mix PCR Reaction Mix kit (iScript™ One-Step RT-PCR Kit with SYBR® Green (Bio-Rad, USA). Thermal cycling conditions were: 10 min at 50°C, 5 min at 95°C then 40 cycles 10 sec at 95°C 30 sec at 55°C, 1 min at 55°C using Rotorgene real time PCR system and the related software for analysis and interpretation (Qiagen- S.Korea). Data were analyzed using the comparative Ct ( $2^{-\Delta\Delta Ct}$ ) method<sup>11,12</sup>. The PCR primers sequences were as follows: \* Casp8 F, 5'-CATCCA GTCACCTTTGCCAGA -3', \* Casp8 R, 5'-GC ATCTGTTTCCCCATGTTT-3', \* AFP F 5' GTCCTTTCTTCCTCCTGGAGAT-3', \* AFP R 5'-CTGTCAGTCT- GATTTCTCTGG-3', \*  $\beta$  -actin was used as a reference gene for internal control, \*  $\beta$ -actin F 5'-GTGACATCC ACACCCAGAGG-3, \*  $\beta$ -actin'R 5'-ACAG GATGTCAAACTGCC-3'.

### Statistical analysis

Data are presented as the mean  $\pm$  SD of three independent experiments. The data were tabulated and analyzed by SPSS (statistical package for the social science software) version 20.0. Statistical significance was analyzed using Student t test and differences were considered statistically significant when P value < 0.05.

### Results

To determine the effect of melatonin on apoptosis, we performed RT-PCR to demonstrate gene expression level of caspases 8, the initiating enzyme for extrinsic pathway of apoptosis with demonstration of the effect of melatonin on AFP gene expression by RT-PCR assay. All experiments are repeated 3 times for more accuracy of the results. Melatonin significantly increased caspase 8 expression. The relative expression was 2.51, 2.48 and 1.89 respectively as compared to control. While the expression of AFP was down regulated by melatonin administration. The relative expression of AFP was 0.051, 0.052 and 0.046 respectively as compared to control. The fold change for caspase 8, AFP is presented in table (1) and figure (1) as mean $\pm$ SD.

Table (1) fold change of Casp 8 and AFP after melatonin and treatment.

| Sample data | Results Fold Change |                   |
|-------------|---------------------|-------------------|
| Sample code | Casp8               | AFP               |
| Melatonin   | 2.29 $\pm$ 0.071    | 0.496 $\pm$ 0.032 |
| Cont.HepG2  | 1                   | 1                 |

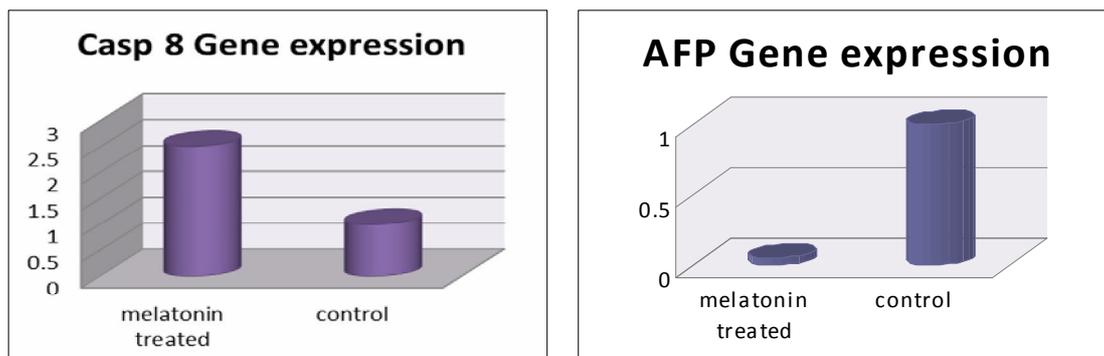


Fig (1) Expression of caspase 8, AFP in melatonin treated groups as compared to non-treated control group showed that it significantly increased caspase 8 expression ( $P < 0.001$ ). Indicating that melatonin promoted apoptosis through initiation of extrinsic pathway, and significantly decreased AFP gene expression ( $P < 0.001$ ).

### Discussion

Surgical resection, local ablation or liver transplantation are possible treatments for a proportion of patients with early stages of HCC. However, these options will be restricted in advanced stages<sup>13</sup>. HCC has limited therapeutic options due to its frequent recurrence, metastasis<sup>14</sup>, and easy relapsing post-operative<sup>15</sup>. Resisting cell apoptosis and sustaining cell growth are two major hallmarks of cancer. During the process of hepatocarcinogenesis, the balance between cell growth and apoptosis will be disrupted<sup>16</sup>. Therefore, selective apoptosis induction in liver cancer cells has emerged as a promising possibility for developing selective cancer therapies<sup>17</sup>. Despite extensive research efforts in this area, there is still controversy concerning the most suitable therapeutic approach to target HCC<sup>18</sup>. Drugs used for treatment of HCC are cytotoxic, and impose a high risk of side effects to the patients. Moreover, it has been recognized that HCC cells are chemo resistant to conventional chemotherapeutic drugs<sup>17</sup>. When sorafenib was firstly tried as a therapy for advanced HCC, it gave a significant survival benefit but unresolved issues are still present Sorafenib is the only presently approved systemic treatment which showed statistically respectable advance in survival and slowed down tumor progression<sup>19</sup>. Recently, there is an increased interest in the potential use of melatonin in clinic<sup>18</sup>. Melatonin is a secretory product of the pineal gland and gastrointestinal tr-

act during night and daytime, respectively<sup>20</sup>. Melatonin is responsible for a variety of physiological functions including antioxidant. Antiapoptotic<sup>21</sup>, and pro-apoptotic activities<sup>16</sup>. So, in this study we aimed to evaluate the effect of melatonin on casp8 mRNA expression and AFP mRNA expression in Hep G2 HCC cell line. According to our study, melatonin decreased AFP expression and stimulates apoptosis via stimulation of casp 8 expression in HCC. Melatonin can be considered a promising adjuvant for chemotherapy in HCC. Previously Shibo et al., 2017. showed that melatonin synergistically enhanced sorafenib-induced apoptosis through induction of caspase-3<sup>22</sup> expression. Based on these data, it is highly suggestive that the combination treatment with both melatonin and sorafenib together may offer better therapeutic potential for patients with HCC. Also in 2008 and in agreement to our results, Renedo et al.<sup>23</sup> showed that melatonin treatment in HepG2 cell line significantly increased casp 8 activity control versus control cell lines. Up till the date no studies known demonstrate the effect of melatonin on AFP gene expression especially at HCC to compare our results with them. It has been reported in clinical trials that supplemental melatonin could abate chemotherapy-related side effects and improve patients' survival by combating several malignant tumors. Melatonin induces apoptosis, promotes cell

cycle arrest, and suppresses angiogenesis and metastasis<sup>16</sup>. Without being toxic to normal cells<sup>24</sup>. Our study introduces melatonin as inhibitor for AFP expression and apoptosis inducer via stimulation of casp 8 expression in HCC. So, Melatonin could be considered a promising adjuvant for chemot-herapy in HCC.

## References

- 1- Holah N, El-Azab D, Aiad H, et al. Hepatocellular carcinoma in Egypt: Epidemiological and histopathological properties. **Menoufia Medical J.** 2015; 28 (3): 718-724
- 2- Kim J, Won R, Ban G et al. Targeted regression of hepatocellular carcinoma by cancer-specific RNA replacement through MicroRNA regulation. **Scientific Reports.** 2015; Article number: 12315.
- 3- Wang X, Wang Q. Alpha-fetoprotein and hepatocellular carcinoma immunity. **Can J. Gastroenterol Hepatol.** 2018; 9049252.
- 4- Tahtouh R, Azzi A, Alaaeddine N et al. Telomerase Inhibition Decreases Alpha-Fetoprotein Expression and Secretion by Hepatocellular Carcinoma Cell Lines: In **Vitro and In Vivo Study.** 2015; PLOS one; 10 (3): e0119512.
- 5- Fulda S. Tumor resistance to apoptosis. **Int J. Cancer.** 2009; 124 (3): 511-515.
- 6- Cosentino K, Garcia-Saez A. Mitochondrial alterations in apoptosis: **Chem Phys Lipids.** 2014; 181: 62-75.
- 7 - Wurstle M, Laussmann M, Rehm M. The central role of initiator caspase-9 in apoptosis signal transduction and the regulation of its activation and activity on the apoptosome. **Exp Cell Res.** 2012; 318 (11): 1213-1220.
- 8- Fulda S. Targeting apoptosis for anticancer therapy. **Semin Cancer Biol.** 2015; 31: 84-88.
- 9- Wu S, Lin W, Shen C, Pan H, Keh-Bin W, Chen Y, et al. Melatonin set out to ER stress signaling thwarts epithelial mesenc-hymal transition and peritoneal dissemina-tion via calpainmediated C/EBPB and NFkB cleavage. **J. Pineal Res.** 2016; 60: 142-154.
- 10- Korkmaz A, Riter R, Topal T, Manchester L, Oter S, Tan D. Melatonin: An established antioxidant worthy of use in clinical trials. **Mol Med.** 2009; 15: 43-50
- 11- Marino J, Cook P, Miller K. Accurate and statistically verified quantification of relative mRNA abundances using SYBR Green I and real-time RT- PCR. **J. Immunol Methods.** 2003; 283: 291-306.
- 12- Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paepe A et al. Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. **Genome Biol.** 3 (7). 2002; RESEARCH0034
- 13- Carbajo-Pescador S, Garcia-Palomo A, Martin-Renedo J, Piva, M, Gonzalez-Gallego, J, Mauriz J. Melatonin modulation of intracellular signaling pathways in hepatocarcinoma HepG2 cell line Role of the MT1 receptor. **J. of Pineal Research.** 2011; 51 (4): 463-471.
- 14- Hu C, Wu J, Cheng C, Wu W. The therapeutic targeting of HGF/c-Met signaling in hepatocellular carcinoma: Alternative approaches. **Cancers.** 2017; 9 (6): 58
- 15- Zhang Y, Li D, Feng F, An L, Hui F, Dang D, Zhao Q. Progressive and prognosis value of notch receptors and ligands in heaptocellular carcinoma: A systematic review and meta-analysis. **Scientific Reports.** 2017; 7 (1): 14809.
- 16- Lin S, Hoffmann K, Gao C, Petrulionis M, Herr I, Schemmer P. Melatonin promotes sorafenib-induced apoptosis through synergistic activation of JNK/c-jun pathway in human hepatocellular carcinoma. **J. of Pineal Research.** 2017; 62 (3): e12398.
- 17- Martin-Renedo J, Mauriz J, Jorquera F, Ruiz-Andres O, Gonzalez P, Gonzalez-Gallego J . Melatonin induces cell cycle arrest and apoptosis in hepatocarcinoma HepG2 cell line. **J. of Pineal Research.** 2008; 45(4): 532-540.
- 18- Carbajo-Pescador S, Martin-Renedo J, Garcia-Palomo A, Tunon M, Mauriz J, Gonzalez-Gallego J. Changes in the expression of melatonin receptors induced by melatonin treatment in hepatocarcinoma HepG2 cells. **J. of Pineal Research.** 2009; 47 (4): 330-338.
- 19- Samonakis D, Kouroumalis E. Systemic treatment for hepatocellular carcinoma: Still unmet expectations. **World J. Hepatol.** 2017; 9 (2): 80-90.

- 20- Margheri M, Pacini N, Tani A, Nosi D, Squecco R, Dama, A et al. Combined effects of melatonin and all-trans retinoic acid and somatostatin on breast cancer cell proliferation and death: Molecular basis for the anticancer effect of these molecules. **European J. of Pharmacology**. 2012; 681 (1-3): 34-43.
- 21- Mortezaee K, Majidpoor J, Daneshi E, Abouzaripour M, Abdi M. Post-treatment of melatonin with CCl4 better reduces fibrogenic and oxidative changes in liver than melatonin co-treatment. **J. of Cellular Biochemistry**. 2017; 119 (2): 1716-1725.
- 22-Shibo L, Hoffmann K, Gao C, Petrulionis M, Herr I, and Schemmer P. Melatonin promotes sorafenib-induced apoptosis through synergistic activation of JNK/c-jun pathway in human hepatocellular carcinoma. **J. of Pineal Research**. 2017; 62(3): e12398.
- 23-Renedo J, Jose´ L, Jorquera F, Ruiz-Andre´s O, González P, González-Gallego J. Melatonin induces cell cycle arrest and apoptosis in hepatocarcinoma HepG2 cell line. **J. Pineal Res**. 2008; 45: 532-540.
- 24- Ordonez R, Carbajo-Pescador S, Prieto-Dominguez N, García-Palomo A, González-Gallego J et al. Inhibition of matrix metalloproteinase-9 and nuclear factor kappa B contribute to melatonin prevention of motility and invasiveness in HepG2 liver cancer cells. **J. Pineal Res**. 2014; 56 (1):20-30