

Original article

A simple bedside blood test (Fibrofast; FIB-5) is superior to FIB-4 index for the differentiation between non-severe and severe fibrosis in patients with chronic Hepatitis C

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Article History	Abstract:
Received: 2/3/2016	A simple noninvasive score (Fibrofast, FIB-5) was developed using
Revised: 30/7/2016	five routine laboratory tests (ALT, AST, Alkaline phosphatase,
Accepted: 1/9/2016	Albumin and Platelets count) for the detection of severe hepatic
	fibrosis in patients with chronic hepatitis C. The FIB-4 index is a
	noninvasive test for the assessment of liver fibrosis, and a score of
	≤ 3.35 enables the correct identification of patients who have non-
	severe (F0-2) from severe fibrosis (F3 4), and could avoid liver
	biopsy. The aim of this study was to compare the performance
	characteristics of FIB-5 and FIB-4 to differentiate between non-
	severe from severe fibrosis. A cross-sectional study included 604
	chronic HCV patients. All liver biopsies were scored using
neywords:	METAVIR system. Both FIB-5 and FIB-4 scores were measured and
Fibrofast (FIB-5)	the performance characteristics were calculated using the ROC
EID A	curve. The performance characteristics of Fibro-Fast at \geq - 2.1 and
1'1 D- 4	FIB-4 at ≤ 3.25 for the differentiation between non-severe fibrosis
Liver fibrosis	and severe fibrosis were; sensitivity 39.6%, NPV 88.7% and
Liver biopsy	sensitivity 29.7%, NPV 87.4% respectively. Conclusion: FIB-5
Hepatitis C Genotype 4	score at the new cutoff is more superior to FIB-4 index for the
I STATEST	differentiation between non- severe and severe fibrosis.

Abbreviations: *ALP*; alkaline phosphatase. *ALT*; alanine aminotransferase. *AST*; aspartate aminotransferase. *CHC*; chronic hepatitis *C*. *HCV*; hepatitis *C* virus. *NPV*: negative predictive value. *PPV*: positive predictive value, *SD*; standard deviation. *Sn*: sensitivity, *SP*: specificity.

1. Introduction

Assessment of liver fibrosis in patients with chronic hepatitis C virus (HCV) infection is considered a relevant part of patient care, for decision making, and is an important prognostic factor [1,2]. In addition, the severity of liver fibrosis may be used as a selection criterion for antiviral therapy, duration of treatment, surveillance for hepatocellular carcinoma and for esophageal varices (HCC), screening [3,4]. The gold standard for assessing the health of the liver is the liver In addition to histological biopsy. evaluation, liver biopsy also provides information on necroinflammatory activity and other features, such as steatosis and hepatic deposits of iron or copper. Besides its advantages, liver biopsy is an invasive technique with associated morbidity, mortality, and complications [5]. Other problems associated with the use of liver biopsy in assessing fibrosis staging are related to sample size and the heterogeneity of pathology in chronic hepatitis C (CHC) infection, and whether this sample is 100% representative of the entire liver [6-12]. Previous reports have proposed that a liver biopsy sample should contain a minimum of 5 portal tracts and be at least 15 mm in length to be considered adequate [8,13-14]. Others have recommended core length >15 mm or contain at least 10 portal tracts [15,16], but this might need more than one path with the biopsy needle to be achieved. Considering these limitations, many studies have recently focused on the development of non-invasive markers as an attractive surrogates of liver biopsy for both patients and physicians [17-27]. Besides the clear advantage of being noninvasive, may overcome the mentioned intra- and interobserver variability of liver biopsy. theoretically these tests offer a more view of fibrogenic events accurate occurring in the entire liver and carry the advantage of providing frequent fibrosis evaluation without additional risk. Fibrofast (FIB-5) and the FIB-4 are a simple noninvasive markers that score for liver fibrosis evaluated on the basis of simple biomarkers (ALT:AST ratio, albumin, alkaline phosphatase, and platelets count) and simple variables such as age, AST, ALT, and platelet count respectively, and can be used by the clinicians to predict severe fibrosis or cirrhosis in patients with CHC infection [27,28]. The objective of this study in Egyptian patients with CHC was to compare the performance characteristics of FIB-5 and FIB-4 to differentiate between non-severe (F0-2) from severe fibrosis (F3-4).

2. Patients and Methods

2.1. Patients

This cross-sectional study conducted on 604 patients chronically infected with HCV genotype 4 (HCV-G4) attending the Egyptian Liver hospital, Mansoura, Egypt in the period from 2013 to 2015. HCV RNA positive patients were identified among HCV antibody (anti-HCV Ab) positive patients. Later, the study plan was discussed with patients and the biopsy was taken only from those patients who were willing for this procedure.

2.2. Patients' Consent

Informed written consent from each patient and local ethical committee

approval were obtained before starting the data collection. With respect to patients' confidentiality, patients were represented in the study by code numbers. All personal data was concealed. The study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the institution's human research committee.

2.3. Exclusion Criteria

Patients who have received any previous courses of antiviral or immunosuppressive therapy, those who had clinical evidence of HBV or HIV infection, and those with any type of liver cancer were excluded from the study. Patients who refused to have a liver biopsy or for whom it was contraindicated, i.e., because of a low platelet count, prolonged prothrombin time or decompensated cirrhosis were also excluded from the study.

2.4. Baseline assessment:

The patients were subjected to thorough history taking, clinical examination, routine laboratory work-up including complete blood count, international normalized ratio (INR), and serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), albumin, and bilirubin.

2.5. Calculations

The FIB-5 score was calculated according to Attallah et al., 2006 [28] as follow:

albumin (g/L)×0.3 + platelet count (10⁹/L)×0.05 - alkaline phosphatase (IU/L)×0.014 + AST/ALT ratio×6 + 14.

The FIB-4 [30] score was calculated as follows:

[Age (year) × AST (IU/L)] / [platelet count (×10⁹/L) × ALT (IU/L)^{1/2}].

2.6. HCV RNA detection and quantitative PCR

RNA was extracted from 140 µl serum samples using OIAamp viral RNA extraction kit (Qiagen USA cat # 52906) according to the manufacturer's protocol. cDNA was synthesized using Moloney murine leukemia virus (MmLV) reverse transcriptase (Invitrogen, USA). First round and nested PCRs were carried out with Taq Polymerase (Fermentas USA) and analyzed on 2% agarose gel. Qiagen HCV quantitative kit was used to perform HCV RNA quantification amplification after each replicating cycle with 10 µl of the extracted RNA on Roche Real Time PCR using fluorescent probes to detect.

2.7. HCV genotyping

HCV genotyping was carried out using Invader HCV genotyping assay (Third wave technology, USA). Briefly, about 100 ng of the HCV RNA was reverse transcribed to cDNA using 200U of MmLV (Invitrogen, USA). From the amplified product, 2 µl was taken and the genotyping assay was performed for 12 different HCV types.

2.8. Liver Biopsy and Histo-pathological examination

The liver biopsy procedure, its advantages, and its possible adverse effects were explained to the patients. An informed consent to obtain a liver biopsy was obtained from all patients. Histopathological examination of ultrasound guided percutaneous liver biopsy requires using 16 gauge semiautomated biopsy needles. Liver specimens of 15mm in length with a minimum of 4 portal tracts were fixed in 10% neutral formalin, then processed and embedded in paraffin. Sections were stained with Hematoxylin and Eosin, and Masson trichrome stains for detection of fibrosis. METAVIR scoring system demonstrated different stages of fibrosis (F0-4) [29]. The histopathological examination of all liver biopsies was performed by a single hepatology expert pathologist. Histological staging based on the degree of fibrosis have five degrees of fibrosis: as F0 (no fibrosis), F1 (mild fibrosis without septa), F2 (moderate fibrosis with few septa), F3 (severe fibrosis with numerous septa without cirrhosis) and F4 (cirrhosis). We further grouped fibrosis stages into two groups; the first as F0-2 (non-severe fibrosis), and the second as F3-4 (severe fibrosis).

2.9. Statistical Analysis

Data were collected in a preformed Data Collection Form, and included demographic, possible mode of HCV infection, clinical, biochemical, serological, and virologic data. All patients' data was tabulated and processed using the Statistical Package of Social Sciences (SPSS) version 15.0 for Windows XP (Chicago, IL, USA). The quantitative data was described with mean, median, standard deviation (SD) or range and compared with student's t-test. Pearson correlation was conducted to correlate continuous parameters. Multivariate backwards stepwise binary logistic regression analysis with severe fibrosis (F \geq 3) - as the dependent factor - was performed. To know how well the FIB-5 test compared to FIB-4 as a diagnostic test can predict that a patient has non-severe (F0-2) or severe fibrosis (F3-4), the statistics positive predictive value (PPV), negative predictive

value (NPV), sensitivity and specificity were used. Efficiency is an overall estimate of a test's ability to classify patients correctly. It is estimated by adding the numbers of the two correct classifications (true positive and true negative) and dividing by the total number of patients assessed. ROC (receiver operator characterristic) curve(s) were constructed to assess area under the curve (AUROC). Patients were classified into two groups (below and above the cutoff values). Best cutoff values for the independent variables were determined based on the nearest point to top left point in the ROC curve. P value <0.05 was considered significant.

3. Results

3.1. Patient's data

According to METAVIR system, the determination of liver fibrosis showed stage F0 in 19 (3.1%), F1 in 372 (61.6%), 112 (18.5%) patients in F2 and 83 (13.7%) patients were F3 stage and 18 (3.0%) patients were F4 (Table 2). According to fibrosis stage, all patients were classified into two groups, the first group considered F0-2 as (non-severe fibrosis) and the second group considered F3-4 as (severe fibrosis).

3.2. Relationship between clinical findings and fibrosis

Liver fibrosis stages were statistically significant between age groups (p=0.000). Non-severe fibrosis was diagnosed mostly in younger patients (<40 years), while more advanced stages were observed in patients over 40 years old, table (1). Liver biochemical tests (serum albumin, platelet count, ALT, AST, and ALP) levels were significantly different in various groups in both groups, table (1). ALT and AST increased in advanced fibrosis, while ALP, albumin, and platelet count decreased in groups of advanced fibrosis. FIB-5 value and FIB-4 differentiated significantly between fibrosis groups (p<0.05), table (1). The relationship between fibrosis stages and the two fibrosis scores (FIB-5 and FIB-4) is illustrated in figure 1 and tables 2 and 3. There was a significant relationship between fibrosis stages and both serum indexes. A significant increase (p=0.000) in the level of FIB-4 as fibrosis progresses from non-severe (F0-2) to severe fibrosis (F3-4). A decrease in the level of FIB-5 observed (p=0.000)was with the progression of fibrosis stages from nonsevere to severe fibrosis, table (1). The AUROCs (P value) of the serum noninvasive indexes are shown in figure (1-a, b B). AUROCs (p value) of FIB-5 for differentiating non-severe fibrosis from severe fibrosis (Figure 1:A) was 0.784 (p=0.000) and for FIB-4 Figure (1:B), it was 0.816 (p=0.000). Table 3 depicts the diagnostic performance of FIB-5 and FIB-4 models for the diagnosis of severe fibrosis stages (F0-2) by using the cutoffs (-2.1, 3.25) for FIB-5 and FIB-4 respectively. When compared to Liver biopsy, FIB-5 values \geq -2.1 showed a NPV of 88.7% for the diagnosis of non-severe fibrosis (F0-2) with sensitivity of 39.6%. On the other hand, FIB-4 at cutoff ≤ 3.25 could indicate to non-severe fibrosis (F0-2) with a sensitivity of 86.5% and a NPV of 87.4%.

Table (1) Baseline characteristics of chronic hepatitis C patients with hepatic fibrosis regards two main classifications (n=604)

Variables	Severe fibrosis	Non severe fibrosis		
	(F3-4)	(F0-2)	p-value	
	(n=101)	(n=503)		
Age (years)	42.2±8.3	36.8±9.6	0.000	
ALT	56(10-435)	42.5(8-239)	0.000	
AST	61(11-264)	40(7-281)	0.000	
ALP	67.5±26.4	73.5±21.1	0.013	
Albumin	4.2±0.4	4.4±0.3	0.000	
Platelet	155.1±63.0	226.1±61.2	0.000	
FIB-4	2.27(0.28-11.70)	1.01(0.11-8.32)	0.000	
FIB-5	-1.10(-26-12.13)	3.9(-18.6-17.5)	0.000	

ALT; alanine aminotransferase. AST; aspartate aminotransferase. ALP; alkaline phosphatase. Data expressed as mean SD or median (range).

Table (2): Frequency distribution of different biopsy groups.

Stage	Frequency	Percent (%)
F0	19	3.1
F1	372	61.6
F2	112	18.5
F3	83	13.7
F4	18	3.0
Total	604	100.0

By METAVIR Score

Table (3) Performance characteristic of both fibro fast and FIB-4 to differentiate between F3, F4 and others fibrosis stages.

	Sn*	Sp	PPV	NPV	Accuracy	No. of detected cases (% of all cases)	No. of false positive cases (% of detected cases)
FIB-5 (-2.1)	39.6%	94.8%	60.6%	88.7%	85.6%	40(6.6%)	26(39.4%)
FIB-4 (3.25)	29.7%	97.8%	73.2%	87.4%	86.5%	30(5%)	11(26.8%)

*Sn: sensitivity, SP: specificity, PPV: positive predictive value, NPV: negative predictive value



Figure (1) Receiver-operating characteristic curves (ROC) curve generated by FIB-5 (A) and FIB-4 (B) for differentiation between severe and non-severe fibrosis. AUROC curve = 0.816 (P = 0.000), 0.784 (P = 0.000) for FIB-4 and FIB-5, respectively.

4. Discussion

For long time, pathological examination of liver puncture tissue was the way to diagnose liver fibrosis. Usage of liver biopsy, because of its invasive trait and sampling errors, is still limited in clinical practice, although it is the gold standard [30,31]. Searching for noninvasive markers to diagnose liver fibrosis has demanded great attention [32-33]. Comparing pathological classification with some noninvasive markers (FIB-5 and FIB-4) to appraise importance of these markers in expressing pathological differences in Egyptian patients with CHC genotype 4 was the main goal of this study. FIB-5 is a newly adopted score that depends on the combination of five routine laboratory markers (albumin, AST, ALT, alkaline phosphatase, and platelet count) for the detection of hepatic fibrosis in patients with CHC [28]. The FIB-4 is also a noninvasive method for the evaluation of liver fibrosis, based on simple variables such as age, AST, ALT and platelet count. It was initially proposed by researchers of the APRICOT study (AIDS Pegasys Ribavirin International Coinfection Trial) to evaluate the presence of liver fibrosis in HIV/HCV coinfected patients [35] and was subsequently validated in HCV monoinfected patients [28]. In the present study, the AUROC of FIB-4 differentiate severe (F3-4, n=101) from non-severe fibrosis (F0-2, n=503) was 0.816, with a sensitivity of 29.7% and a specificity of 97.8% for cutoff of 3.25. The NPV was 87.4 % and the PPV was 73.2% with an accuracy of 86.5%. Similar results were obtained by other authors who evaluated the performance of FIB-4 in HCV monoinfected patients, with the

AUROCs ranging between 0.732 and 0.799 [36-40]. When the diagnostic performance of FIB-5 was compared to the FIB-4, FIB-5 was found to be better than FIB-4 for diagnosing non-severe (F0-2). FIB-5 using the new cutoff value (> -2.1) showed a NPV of 88.7% for the diagnosis of non-severe fibrosis (F0-2) with sensitivity of 39.6%. According to the results of Attallah et al., 2006 [28], a cutoff zero on FIB-5 score was previously suggested to be a cutoff point between F0-2 and F3-4 on METAVIR staging with 98% sensitivity, 97% specificity, 99% PPV, and 92% NPV. Fenili Amorim et al., 2012 [42] observed an AUROC of FIB-4 to detect significant fibrosis of 0.811±0.045, with a sensitivity of 63% and 28% and a specificity of 82% and 99% for cutoffs of <1.45 and >3.25 respectively. The NPV was 81% for FIB-4 values. Vallet Pichard et al [30] observed an AUROC of 0.85 for identifying fibrosis (F3-4), In HCV monoinfected patients. The cutoff point <1.45 showed a NPV of 94.7%, with a sensitivity and specificity of 74.3% and 80.1% respectively. FIB-4 values 3.25 have a PPV of 82.1% with lower sensitivity (37.6%) and higher specificity (98%). In the study of Elnakeeb et al., 2014 [43], the FIB-4 index proved to be sensitive and specific in the differentiation between patient with no or mild fibrosis (METAVIR F0-1) and patients with significant fibrosis or cirrhosis (F2-4) (AUC=91.6) with the best cutoff value at 1.61 where sensitivity was 69.5% and specificity was 100%. The PPV was 100% to detect patient with no or mild fibrosis. Using this cutoff (1.61), 87% of patients fell outside these ranges and could

thus avoid liver biopsy with an overall accuracy of 70%. Sterling et al., 2006 [36] who proposed the use of FIB-4 index in patients with HIV/HCV coinfection, found that FIB-4 index can differentiate between Ishak stage 0-3 and 4-6. At a cutoff of <1.45, the negative predictive value to exclude advanced fibrosis (stage 4-6) was 90% with a sensitivity of 70%. A cutoff of >3.25 had a positive predictive value of 65% and a specificity of 97%. Vallet-Pichard et al., 2007 [30] evaluated the use of FIB-4 index in 847 patients with HCV monoinfection, found that FIB-4 index higher than 3.25 had a positive predictive value to confirm the existence of a severe fibrosis (F3-4) of 82.1% with a specificity of 98.2%. Sumida et al., 2012 [44] evaluated the use of FIB-4 in 576 patients with nonalcoholic fatty liver disease, found that a FIB-4 index higher than 3.25 had a positive predictive value to confirm the existence of a severe fibrosis.

5. Conclusion

This study demonstrated that FIB-5 was superior to FIB-4 in the diagnosis of nonsevere fibrosis in patients with chronic hepatitis c.

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