Prevalence of occult HCV infections in Saudi patients who achieved sustained virologic response with direct acting antiviral treatment

Hamad Al Ashgar¹, Ahmed AlQuahtani², Mohammed Al Quaiz¹, Hussa Alhussaini³, Khalid Al Kahtani¹, Amr M Elssadany⁴, Saleh A Alqahtani⁵, Musthafa Chalikandy Peedikayil*¹.

¹Medicine, Gastroenterology dept., King Faisal Specialist Hospital & Research Center, Riyadh, Saudi Arabia, ²Infection and Immunity dept., Research Center, King Faisal Specialist Hospital & Research Center, Riyadh, Saudi Arabia, ³Anatomic pathology, King Faisal Specialist Hospital & Research Center, Riyadh, Saudi Arabia, ⁴Radiology dept., King Faisal Specialist Hospital & Research Center, Riyadh, Saudi Arabia, ⁵Consultant hepatologist, King Faisal Specialist Hospital and Research Centre. Riyadh, Saudi Arabia.

Received: 17-6-2020 Accepted: 13-8-2020

Abstract

Background: Occult hepatitis C virus (HCV) infection (OCI) is a condition where HCV RNA is present in hepatocytes or peripheral blood mononuclear cells (PBMCs), but not in the serum, in patients with active or past HCV infection. Serum HCV antibodies may or may not be present. Aim: We investigated the prevalence of OCI in PBMCs and hepatocytes of patients who had achieved sustained virologic response (SVR) after 24 weeks of direct-acting antiviral treatment. Methods: Chronic HCV patients with Genotype 1a and 4 who achieved SVR24 weeks after treatment with direct-acting antiviral agents (DAAs) were prospectively selected. RNeasy Blood and Tissue Kit (Qiagen GmbH, Germany) was used for RNA extraction from blood and liver tissue samples. Superscript IV First-Strand Synthesis System (Invitrogen) was used for reverse transcription PCR. Quantitative and qualitative detection of HCV RNA was performed using primers specific to the 5'untranslated region (5'UTR). Patients: Of the six chronic HCV patients recruited for this study, five were infected with genotype 4 and one with genotype 1a. Five patients were treated with Sofosbuvir and Daclatasvir, and one patient with Ledipasvir plus Sofosbuvir. All of the patients were immunocompetent. Results: None of the patients had detectable HCV RNA in either the PBMCs or hepatocytes, suggesting zero prevalence of OCI in patients who achieved 24 weeks SVR post DAA treatment. Conclusion: We failed to detect HCV RNA in both the hepatocytes and PBMCs of all patients recruited for the study. This finding suggests that OCI is a rare phenomenon in chronic HCV patients with SVR following DAAs.

Keywords: Chronic hepatitis, HCV, hepatitis C virus, occult infection, occult HCV infection, direct-acting antiviral agents.
OCI was reported in the era of interferon and ribavirin treatment, its clinical relevance after the introduction of DAA has not been thoroughly investigated. Recent studies on explanted livers following liver transplantation demonstrated the prevalence of OCI in patients treated with DAA after having SVR. Another study reported the presence of OCI in PBMCs after achieving SVR post DAA treatment. These studies show the varying occurrence of OCI in patients who have achieved SVR post DAA treatment. In Saudi Arabia, a national program for HCV elimination was launched last year. The objective of the program was to implement efficient treatment methods to cure HCV infections in patients from Saudi Arabia; thus, it is critical to know if OCI exists in patients who achieved SVR after treatment. To our knowledge, there is no data available or published work addressing OCI in Saudi patients. The current study is a pilot analysis to study OCI in chronic HCV patients from Saudi Arabia, who achieved SVR of 24 weeks post-treatment with DAA.

Materials and Methods

Study design
This was a descriptive cross-sectional survey.

Study Area
The study was performed in patients from Saudi Arabia, who underwent treatment at the HCV clinic at King Faisal Specialist Hospital and Research Center, Riyadh, between 15th October 2017 and 11th March 2018. Hepatologists conducted the study in collaboration with the Department of Research, the Department of Pathology, and the Department of Radiology at the King Faisal Specialist Hospital and Research Center. The research promotion committee at the Department of Medicine and Department of Research Advisory Council and Ethics at the King Faisal Specialist hospital approved the study in July 2017 (RAC number #2171098). All patients included in the research consented to be studied and were counseled accordingly.

Study population
All patients recruited for this study had prior chronic HCV infection and were treated with DAA, who subsequently achieved SVR at 12 and 24 weeks. No HCV RNA was detected in blood samples at 12- and 24-weeks post-DAA treatment. The patients had a fibrosis score of F1 to F3 on fibroscan. Pregnant, lactating women, patients with thrombocytopenia (values of less than 50,000 platelets/µl of blood and coagulopathy with international normalized ratio (INR) >1.5), patients who had a infection of both HCV and HIV, and patients who had recent organ transplantation were not included in the study.

Specimen collection and isolation of PBMCs from blood samples
After collecting the relevant information and explaining to the patients the aim of the study and the test procedures, blood samples were collected aseptically in tubes containing ethylene diamine tetra acetic acid (EDTA). PBMCs were isolated following standardized protocols.

Liver biopsy
The patients underwent transcutaneous liver biopsy guided by ultrasound done by a licensed radiologist and following the accepted guidelines, after obtaining consent from the patients. The length of liver tissue harvested from the liver biopsy was ~5 mm and was performed using a Tru-Cut needle biopsy. Two tissue specimens were harvested from each patient, one collected in formalin solution for analysis at the pathology department and another in RNA stabilization reagent (Qiagen GmbH, Germany) for HCV RNA detection at the research department.

RT-PCR and RNA isolation from PBMCs and hepatocytes
RNA was extracted and purified from PBMCs and hepatocytes (harvested from liver tissues following the standardized protocol) using RNeasy Blood and Tissue Kit (Qiagen GmbH, Germany). Reverse transcription PCR (RT-PCR) of the RNA extracted from the samples was done following previously standardized protocols. RT-PCR of the RNA was performed using the superscript IV first-strand synthesis system (Invitrogen Life Technologies, Carlsbad CA, USA) to synthesize complementary DNA (cDNA), and quantitative and qualitative detection of HCV RNA PCR amplification was performed using primers specific to the 5’ untranslated region (5’UTR). A product of 241bp was formed representing the HCV RNA.

Sample size
The current analysis is a pilot study to test the feasibility of testing for OCI by detecting the presence of HCV-RNA in hepatocytes and PBMCs using RT-PCR. Six patients were recruited for this study and provided consent for liver biopsy.

Data entry
All eligible patients were clinically assessed throughout the study period by having procedural blood tests, imaging studies, and fibroscan. The data was documented cautiously following protocols. Data from the DAA regimen, HCV RNA viral load of blood samples by RT-PCR, HCV genotype, liver biopsy with the grading of inflammation and staging of fibrosis (Metavir score), and the results of HCV RNA by RT-PCR from PBMCs and hepatocytes were also cautiously documented.
Statistical data analysis

Based on the data acquired, categorical variables were analyzed using numbers, ratios, and frequency tables. For discrete variables and continuous variables, mean, median, range, and standard deviation were analyzed. A Chi-square test was used to study the relationship between two categorical variables. When one or more cells had an expected frequency of five or less, Fischer’s exact test was used for the analysis. For two independent groups, an independent samples t-test was used to compare the mean of normally distributed interval dependent variables.

Results

A total of six patients were recruited for this study, out of which five were males, and one was a female with a mean age of 42.3 years. The data from peripheral blood RT-PCR obtained from all six patients, and hepatocytes RT-PCR obtained from five patients was analyzed. Liver function tests performed during liver biopsy had a mean alanine aminotransferase (ALT) value of 16.8 IU/ml (ALT in the recruited patients were normal at the time of liver biopsy). Five patients were treated with a combination of daclatasvir and sofosbuvir, and one patient was treated with sofosbuvir and ledipasvir. Five patients were genotype 4, and one was infected with genotype 1a. Baseline characteristics of the recruited patients who underwent either liver biopsy or blood tests are shown in tab. (1). None of the patients showed any complications due to liver biopsy. Each patient’s HCV viral load before the initiation of DAA treatment and at the time of liver biopsy are given in the table with title baseline characteristics. The liver biopsy findings were assessed by Metavir scoring system; histology of the liver tissue showed fibrosis ranging from stage I to stage III; inflammatory activity in the liver biopsy ranged from grade 0 to grade 3. HCV viral load in patients was in the range from 0.37 million IU/ml copies to 43.3 million IU/ml copies. RT-PCR results of PBMCs and hepatocytes isolated from patients showed no presence of HCV RNA in any of the samples, fig. (1 & 2). The prevalence of OCI among the recruited patients was zero. Further statistical calculations were not performed because of a lack of evidence of OCI prevalence.

Table 1. Characteristics of the patients who were involved in the study.

<table>
<thead>
<tr>
<th>Case</th>
<th>Post-Treatment Liver biopsy</th>
<th>Post-Treatment blood sample tested</th>
<th>HCV Q before the DAA treatment</th>
<th>HCV Q before the Liver biopsy</th>
<th>Liver biopsy grading and staging</th>
<th>Genotype</th>
<th>Direct-acting antiviral agents</th>
<th>HCV from WBC by RT PCR</th>
<th>HCV from the liver tissue by RT PCR</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Done</td>
<td>Done</td>
<td>409179</td>
<td>0</td>
<td>F0, S1</td>
<td>4</td>
<td>Sofosbuvir and Daclatasvir</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>2</td>
<td>Done</td>
<td>Done</td>
<td>43318145</td>
<td>0</td>
<td>F1, S1</td>
<td>4</td>
<td>Sofosbuvir and Daclatasvir</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>3</td>
<td>Done</td>
<td>Done</td>
<td>2131441</td>
<td>0</td>
<td>F1, S1</td>
<td>4</td>
<td>Sofosbuvir and Daclatasvir</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>4</td>
<td>Done</td>
<td>Done</td>
<td>373520</td>
<td>0</td>
<td>F0, S1</td>
<td>4</td>
<td>Sofosbuvir and Daclatasvir</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>5</td>
<td>Done</td>
<td>Done</td>
<td>4529417</td>
<td>0</td>
<td>F3, S3</td>
<td>1a</td>
<td>Ledipasvir plus Sofosbuvir</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>6</td>
<td>Not done</td>
<td>Done</td>
<td>373467</td>
<td>0</td>
<td>Liver biopsy not done</td>
<td>4</td>
<td>Sofosbuvir and Daclatasvir</td>
<td>Negative</td>
<td>Not done</td>
</tr>
</tbody>
</table>

Figure 1. HCV RNA detected by reverse transcription PCR were negative in the hepatocytes of all five tested patients. Lane 1, DNA molecular weight; lane 2, HCV RNA positive control; lanes 3-8, patient samples.
Discussion

Occult HCV infection (OCI) is a relatively new phenomenon identified in the last decade\(^8\). OCI is described as a condition where HCV RNA is detected in the hepatocytes or PBMCs, but not in the serum, with or without serum HCV antibodies. The current analysis was done to investigate the prevalence of OCI in both PBMCs and hepatocytes in chronic HCV patients treated with DAAs who achieved SVR at 24 weeks post-treatment. We found that, post-DAA treatment 24 weeks SRV, none of the patients had evidence of OCI in either PBMCs or hepatocytes. At present, OCI is observed in three different clinical settings: serum negative HCV RNA and HCV antibodies with consistently elevated liver enzymes and detectable HCV RNA in the liver tissue or PBMCs; serum negative HCV RNA and serum positive HCV antibodies with normal liver enzymes and detectable HCV RNA in the liver tissue or PBMCs; serum negative HCV RNA and serum positive HCV antibodies with abnormal liver enzymes and detectable HCV RNA in the liver tissue or PBMCs\(^4\).\(^5\).\(^6\).\(^7\).\(^8\).\(^9\).\(^10\).

Multiple studies have suggested that HCV RNA in PBMCs might act as an extrahepatic site for HCV replication\(^5\).\(^7\). Also, study findings support the notion that OCI is a potential culprit in cryptogenic liver disease. OCI in cryptogenic liver cirrhosis has been reported to range between 8.9\% and 40\% in PBMCs and between 0\% and 50\% in hepatocytes\(^5\).\(^8\).\(^9\).\(^10\). During the period when interferons, including pegylated interferon and ribavirin, were used to treat chronic hepatitis C infections, several studies reported the presence of viable HCV genome in PBMCs and hepatocytes in patients, many years after treatment. The patients were considered to be clinically cured of HCV infection, confirmed by the absence of HCV RNA in the serum\(^4\).\(^8\).\(^9\).\(^10\). Extremely low levels of viral particles, levels under the detection limit of conventional RT-PCR, could account for why no HCV RNA was detected in the serum of patients who had viable virus particles in the liver and PBMCs\(^5\).\(^7\). A study by Gamabato et al. analyzed HCV RNA in the liver explants of 39 patients treated with an interferon-free regimen\(^\footnote{9}\), with only 6 (15\%) patients finalized for the treatment course at the time of liver transplantation. HCV RNA was detected and quantified in 26/39 liver explants (67\%), and most patients in the cohort were treated with sofosbuvir and ribavirin. The presence of HCV RNA in the liver explants showed no association with virologic relapse after transplantation, except in patients with elevated levels of HCV RNA\(^\footnote{9}\). A study similar to the current analysis, by Yousif et al., on the prevalence of OCI in PBMCs in patients who were treated with DAA with an SRV of 12–24 weeks post-DAA treatment, reported an OCI rate of 11.33\% in PBMCs. However, none of the patients underwent a liver biopsy to detect OCI in their hepatocytes\(^\footnote{10}\). Another study by Elmasry et al. identified a cohort of 14 post-liver transplant patients who were treated with DAA and achieved SVR. However, the levels of serum aminotransferases failed to normalize during or after DAA therapy or normalize transiently but elevated rapidly post-DAA therapy. Nine out of 14 patients had a liver biopsy and were tested for OCI by RT PCR. Out of the nine patients, five patients (55\%) were found to have OCI infection\(^\footnote{5}\). Wang et al, from China studied 140 subjects, and from them 16 had features of occult HCV (11.4\%) and 60 patients from 140 had treatment with DAA, and 9 (15\%) of them showed features of occult HCV\(^\footnote{11}\). A study from Iran, 161 intravenous drug users with HIV infection were enrolled. These subjects were screened for HCV infection and 27 patients were found anti-HCV negative in the serum and negative HCV RNA in the plasma, and from these 27 subjects 5 (18.5\%) of the patients were found to have occult HCV from the PBMCs. OCI was tested by looking for HCV RNA in PBMCs\(^\footnote{12}\). Another study from Iran enrolled HIV-infected 190 Iranian individuals, and they were tested for occult HBV and occult HCV by doing Viral RNA.

Figure 2. HCV RNA detected by reverse transcription PCR were negative in the peripheral blood mononuclear cells of all six patients. Lane 1, DNA molecular weight; lane 2, HCV RNA positive control; lanes 3-10, patient samples.
and DNA extraction from plasma and PBMC specimens, and the presence of HCV-RNA in plasma and PBMC samples was tested using reverse transcriptase-nested polymerase chain reaction (PCR), HBV viral load was determined in plasma samples using COBAS TaqMan 48 Kit, and also the presence of the HBV-DNA in PBMC samples was tested by real-time PCR. In this study, the prevalence of OBI and OCI in HIV-infected individuals was 3.1% and 11.4%, respectively. Contrary to the results from these two studies, our study showed no patients to have OCI infection with no detectable HCV RNA in either the PBMCs or liver tissue after achieving SVR of 12-24 weeks post-DAA treatment tab. (3). It is critical to note that all patients in this study were immune-competent, in contrast to the post-liver transplant patients studied by Elmasry et al. Also, the study by Yousif et al. analyzed OCI only in PBMCs, and none of the patients underwent a liver biopsy to detect OCI in the liver tissue. The clinical significance of OCI observed by Yousif et al. and Elmasry et al. are still unknown. Based on the results reported by Elmasry et al., OCI testing can be recommended to patients who have SVR with persistently elevated aminotransferases or fluctuating aminotransferases, but further investigation is warranted to validate this. A limitation of the current study is its small sample size, with only six patients in total. However, the fundamental approach of this investigation was to understand the concept of occult HCV in a Saudi population analyzed in both PBMCs and liver tissue. Further research is required in a more extensive group of patients, and further validation in the future could help develop screening tests for occult HCV in individuals who have SVR and are potential organ and blood donors and for premarital HCV screening. Also, concordance analysis of OCI in both hepatocytes and PMBCs needs further investigation. If a concordance value close to 100% is achieved, OCI could be tested for less invasively using only PMBCs, thus avoiding invasive liver biopsy procedures. Here, all six patients tested negative for OCI in both blood and liver tissue.

Table 2. Differences between the three types of occult HCV.

<table>
<thead>
<tr>
<th>Occult HCV Types</th>
<th>HCV RNA from liver tissue or PBMCs</th>
<th>HCV Antibody</th>
<th>Liver enzymes</th>
<th>HCV RNA</th>
<th>Occult HCV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type 1</td>
<td>Negative</td>
<td>Negative</td>
<td>Abnormal</td>
<td>Positive</td>
<td></td>
</tr>
<tr>
<td>Type 2</td>
<td>Negative</td>
<td>Positive</td>
<td>Normal</td>
<td>Positive</td>
<td></td>
</tr>
<tr>
<td>Type 3</td>
<td>Negative</td>
<td>Positive</td>
<td>Abnormal</td>
<td>Positive</td>
<td></td>
</tr>
</tbody>
</table>

Table 3. Summary of previous studies on Occult HCV infection in DAA-treated patients with reverse transcription PCR for HCV RNA from hepatocytes.

<table>
<thead>
<tr>
<th>Trial</th>
<th>Characteristics of patients</th>
<th>Liver biopsy</th>
<th>Occult HCV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Elmasry⁸</td>
<td>14 patients treated with DAA with abnormal LFT after SVR</td>
<td>9/14 patients had liver biopsy</td>
<td>6/9 (55%)</td>
</tr>
<tr>
<td>Gambato⁹</td>
<td>• Rx Duration 14 weeks¹⁰-²²</td>
<td>39 patients explanted liver</td>
<td>26/39 positive HCV in the liver explant.</td>
</tr>
<tr>
<td></td>
<td>• Duration from undetectable HCV RNA in the serum to Liver Transplant = 61 days (15–118).</td>
<td></td>
<td>6/9 positive in the DAA group</td>
</tr>
<tr>
<td>The current study from Saudi Arabia</td>
<td>6 chronic HCV patients with F1 to F3 fibrosis, with SVR 24 weeks after DAA treatment</td>
<td>5/6 patients</td>
<td>None had occult HCV</td>
</tr>
</tbody>
</table>

Conclusion
In conclusion, results from the current pilot study showed no evidence of OCI in patients from Saudi Arabia in both hepatocytes and PBMCs, suggesting that occult HCV is a rare phenomenon, especially among immunocompetent patients post-DAA treatment with SVR of 24 weeks. A prospective investigation in a large sample size is warranted, which could further help validate testing for OCI in the future.
References